International Journal of Current Advanced Research

ISSN: O: 2319-6475, ISSN: P: 2319-6505, Impact Factor: 6.614 Available Online at www.journalijcar.org Volume 7; Issue 8(B); August 2018; Page No. 14654-14657 DOI: http://dx.doi.org/10.24327/ijcar.2018.14657.2667



PREVALANCE OF SELECTED SEROTYPE OF ENTEROBACTERIACEAE PATHOGENS FROM SEWAGE AND DRINKING WATER ENVIRONMENT

Archana G¹., Judia Harriet Sumathy V² and Balaji M³

^{1,2}Department of Biotechnology, Women's Christian College, Chennai, India ³Department of Bioinformatics, ABS-Geno-Informatics, Chennai, India

ABSTRACT ARTICLE INFO Enterobacteriaceae are a large family of bacteria that include pathogens such as Salmonella Article History: and Escherichia coli. They are facultative anaerobes which ferment sugar to produce lactic Received 04th May, 2018 acid and are found in water and soil. E.coli of serotype O157:H7 causes food poisoning in Received in revised form 16th humans and serious illness. It has the ability to transfer DNA via conjugation, June, 2018 Accepted 25th July, 2018 transformation and allow genetic material to spread in existing population. Virulent strains Published online 28th August, 2018 of E.coli cause gastroenteritis, urinary tract infection and neonatal meningitis. Examples include Entero toxigenic E.coli which use fimbrial adhesins to bind to enterocyte cell in Key words: small intestine and produce LT and ST enterotoxin. On the other hand Entero pathogenic

Enterobacteriaceae, Escherichia coli, Serotype, Sewage and Drinking Water Environments

E.coli which lack fimbriae use adhesin to bind to intestinal cells which cause significant deformation. Entero invasive E.coli in humans use adhesin protein to bind and enter intestinal cells but do not produce toxin but damage cell. The present study is aimed at studying the Serotype of Enterobacteriaceae pathogens isolated from sewage and drinking water environments.

Copyright©2018 Archana G., Judia Harriet Sumathy V and Balaji M. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

E. coli is a type of fecal coliform bacteria commonly found in the intestines of animals and humans (Acheson et.al., 1998). The presence of E. coli in water is a strong indication of recent sewage or animal waste contamination (Ahmed et.al., 2000). Sewage may contain many types of disease-causing organisms. Fecal coliforms are bacteria that are associated with human or animal waste (Bach et.al., 2002). Presence of fecal coliforms in water may not be directly harmful, and does not necessarily indicate the presence of feces, however it does indicate an increased likelihood of harmful pathogens in the water (Beutin et.al., 1993). Escherichia coli (abbreviated as E. coli) are bacteria found in the environment, foods, and intestines of people and animals. E. coli are a large and diverse group of bacteria (Chalmers et.al., 2000). Although most strains of E. coli are harmless, others can make you sick (Gascon et.al., 1998). Some kinds of E. coli can cause diarrhea, while others cause urinary tract infections, respiratory illness and pneumonia, and other illnesses (Ingledew, W.J. and Poole, R.K. 1984).

E. coli O157:H7 is most commonly found on a small number of cattle farms where the bacteria can live in the intestines of healthy cattle (Meadand, P.S. and Griffin, P.M. (1998).

*Corresponding author: Archana G Department of Biotechnology, Women's Christian College, Chennai, India

Millions of germs can be released in a bowel movement from an infected human or animal. E. coli may also be found in water sources, such as private wells, that have been contaminated with feces from infected humans or animals (Muniesa et.al., 2006). Waste can enter the water through different ways, including sewage overflows, sewage systems that are not working properly, polluted storm water runoff, and agricultural runoff (Rahn et.al., 1998). Wells may be more vulnerable to such contamination after flooding, particularly if the wells are shallow, have been dug or bored, or have been submerged by floodwater for long periods of time (Tarr et. al., 2002). Most strains are harmless and live in the intestines of healthy humans and animals. However, this strain, O157:H7, produces a powerful toxin that can cause severe illness (Varma et.al., 2003). The present study is aimed at studying the Serotype of Enterobacteriaceae pathogens isolated from such sewage and drinking water environments.

MATERIALS AND METHODOLOGY

By plate count method 1 ml of the sample was prepared and transferred to 9 ml of saline and was maintained as master dilution. From this $(10^{-1} \text{ to } 10^{-6})$ dilutions were prepared and 1 ml of sample was poured to cool sterilized agar count plate and incubated at 37°C for 24 hours. Colony was counted by colony counter. Morphological study was achieved by microscopic observation of Grams staining, Motility test, Catalase test and Oxidase test. A small portion of suspected colony was streaked on medias such as Nutrient Agar, MacConkey Agar and Eosin Methylene Blue Agar. Biochemical tests were performed using Standard Protocol. Following this serological typing was done. Depression plates were taken and were marked as A, B and C. In A depression plate it was marked as negative control in which phenolized saline suspension was added. In B depression plate it was marked as test in which phenolized saline suspension and antiserum of respective organism was added and in C depression plate it was marked as positive control which contain phenolized saline suspension of known organism and antiserum.

RESULTS AND DISCUSSION

The total number of positive and negative samples obtained from sewage and drinking water for *E.coli* was found to be 30. In identification of bacterial isolate of morphological characteristics by Grams staining and motility for *E.coli*, it was found to be Gram negative small rods and motile. Cultural characteristics of *E.coli* on Nutrient Agar formed small colonies whereas on MacConkey Agar formed pink colour non mucoid colonies and on Blood Agar Non haemolytic colonies. Biochemical Test, Antibiotic Sensitivity Test and Serotype study results indicate the prevalence of *E. coli* to be dominant in the Sewage and Drinking water samples collected (Tables 1 – 4 and Figures 1 - 5).

Table 1 Biochemical Tests for E.coli

S.No.	Biochemical Test	Result
1.	Catalase	Positive
2.	Oxidase	Negative
3.	Triple Sugar	Acid
4.	Iron Agar	Positive
5.	Gas	Positive
6.	Hydrogen Sulphate	Positive
7.	Indole	Positive
8.	Methyl Red	Positive
9.	Voges Proskauer	Negative
10.	Citrate	Negative

Table 2 Antibiotic Sensitivity Test for E.coli

S.No.	Antibiotics	Zone of Inhibition (mm)
1.	Amikacin	18
2.	Chloramphenicol	22
3.	Co-trimoxazole	22
4.	Tetracycline	13
5.	Gentamycin	17
6.	Ceftriaxone	30
7.	Cephotaxime	19
8.	Norfloxacin	24
9.	Meropenem	10
10.	Imipenem	25

Zone of inhibition

- Below 10mm least active Between 11-25mm – active
- Above 26mm very active

 Table 3 Serotypying of E.coli from Sewage Sample

S. No.	Antiserum	Isolates	Result
1.	O157:H7	1,2,3,6,7,8,10,12,13,14,16,17,20,24,25,26,27,28	Positive
		4,5,9,11,15,18,19,21,23,29,30	Negative
2.	O104:H21	4,5,9,11,15,18,19,21,23,29,30	Positive
		1,2,3,6,7,8,10,12,13,14,16,17,20,22,24,25,26,27,28	Negative

 Table 4 Serotypying of E.coli from Drinking Water Sample

S. No.	Antiserum	Isolates	Result
1.	O157:H7	1,2,3,4,5,6,7	positive
		6,7	negative
2.	O104:H21	6,7	positive
		1,2,3,4,5	negative



Figure 1 Antibacterial activity of E.coli







Figure 3 Serotyping of E.coli from water sample



Gram Negative Rods Catalase Test



Positive Negative

Oxidase Test



A = Negative Control B = Positive Control C = Test Sample









A - *E.coli* on Nutrient Agar B - *E.coli* on MacConkey Agar C - *E.coli* on Blood Agar D - *E.coli* on EMB Agar



A = Indole B = Methyl Red C = Voges Proskaur D = Citrate E = TSI



Figure 5 Antibiotic sensitivity test for *E.coli*

Environmental pollution by faecal contamination is a major threat to the community, which was highly ignored in the past. Drinking water can become contaminated with human or animal faeces by surface run-off and septic tank malfunction etc. When faecal contamination occurs, there is potential for disease causing germs to be present. Similar to ozone problem, green house effect and increase of atmospheric heat, this problem should also be dealt in seriously not only by environmentalists but also by every common man. It may be a true dream that India can emerge to be among the world's first four economic powers by 2020. Unless otherwise environmental pollution by faecal contamination is looked after genuinely, India cannot become a developed country in 2020.

References

- Acheson, D.W., Frankson, K., Willis, D. and STEC (1998). Prevalence Study Group Multice Enterobactericeae prevalence study of Shiga toxin-producing *Escherichia coli*. Abstracts of the 98th General Meeting for the American Society for Microbiology Washington American Society for Microbiology.
- Ahmed, A., Li, J., Shiloach, Y., Robbins, J. and Szu, S. (2006). Safety and immunogenicity of *Escherichia coli* O157 O-specific polysaccharide conjugate vaccine in 2-5-year-old children *J*, *Infectious Disease* 193(4):515-21.
- Bach, S.J., McAllister, T.A., Veira, D.M., Gannon, V.P.J. and Holley, R.A. (2002). Transmission and control of *Escherichia coli* O157:H7. *Canadian Journal of Animal Science* 82:475-490.
- Beutin, L., Geier, D., Steinruck, H., Zimmermann, S. and Scheutz, F. (1993). Prevalence and some properties of verotoxin (Shiga-like toxin)-producing *Escherichia coli* in seven different species of healthy domestic animals. *Journal of Clinical Microbiology* 31, 2483-248
- Chalmers, R.M., Aird, H. and Bolton, F.J. (2000). Waterborne *Escherichia coli* O157. Symposium Series Society for Applied Microbiology 88, 124S–132S.
- Gascon, J., Vargas, M., Quinto, L., Corachon, M., Jimenez de Anta, M.T. and Vila, J. (1998). Enteroaggregative *Escherichia coli* strains as a cause of traveler's diahorrea a case-control study. *J. Infectious Disease* 177:1409–1412.

- Ingledew, W.J. and Poole, R.K. (1984). The respiratory chains of *Escherichia coli*. Microbiology Review 48(3): 222-71. PMID 6387427.
- Lior, H. and Gyles, C.L. (1994). Classification of *Escherichia coli*, in *Escherichia coli* in domestic animals and humans. Wallingford: *CAB International*; 31-72.
- Meadand, P.S. and Griffin, P.M. (1998). *Escherichia coli* O157:H7, *Lancet* 352:1207-1212.
- Muniesa, M., Jofre, J., Garcia-Aljaro, C. and Blanch, A.R. (2006). Occurrence of *Escherichia coli* O157:H7 and other Enterohaemorrhagic *Escherichia coli* in the environment. *Environment Science Technology* 40, 7141-7149.
- Rahn, K., Renwick, S.A., Johnson, R.P., Wilson, J.B., Clarke, R.C., Alves, D., McEwen, S.A., Lior, H. and Spikam, J. (1998). Follow-up study of verocytotoxigenic *Escherichia coli* infection in dairy farm families. *Journal of Infectious Disease* 177 (4):1139-1140. PMID 9535003.
- Tarr, C.L., Large, T.M., Moeller, C.L., Lacher, D.W., Tarr, P.I., Acheson, D.W. and Whittam, T.S. (2002). Molecular characterization of a serotype O121:H19 clone, a distinct Shiga toxin-producing clone of pathogenic *Escherichia coli*. *Infectious Immunology*; 70:6853-9.
- Varma, J.K., Greene, K.D., Reller, M.E., Delong, S.M., Trottier, J., Nowicki, S.F., DiOrio, M., Koch, E.M., Bannerman, T.L., York, S.T., Lambert-Fair, M.A., Wells, J.G. and Mead P.S. (2003). An outbreak of *Escherichia coli* O157 infection following exposure to a contaminated building. *JAMA* 290(20): 2709-2712. PMID 14645313.

How to cite this article:

Archana G *et al* (2018) 'Prevalance of Selected Serotype of Enterobacteriaceae Pathogens From Sewage and Drinking Water Environment', *International Journal of Current Advanced Research*, 07(8), pp. 14654-14657. DOI: http://dx.doi.org/10.24327/ijcar.2018.14657.2667
